



Detection of gastrointestinal nematodes resistance against Ivermectin in sheep

N. Kh. Ibrahim¹, S. D. Hassan², and Q. T. Al-Obaidi²✉

¹Department of anesthesia, Medical Technical Institute, Northern Technical University, Mosul, Iraq.

²Department of Internal and Preventive Medicine, College of Veterinary Medicine, University of Mosul, Mosul, Iraq

Submitted: August 18, 2025

Revised: August 24, 2025

Accepted: August 29, 2025

Abstract To date, Ivermectin is one of the advisable medications widely used in the therapy, domination effort of internal and external parasites in ruminants, but its efficacy and resistance developing during the last years remain questionable. The efficacy of Ivermectin and the resistance state of gastrointestinal; round worms in sheep farms was the goal of the current work survey. 300 fecal samples collected from male sheep (≥ 7 months) represented 6 sheep flocks ($n=730$) from different regions in Mosul city, Iraq. The efficacy of the Ivermectin in one farm with EPG ≥ 200 was evaluated using a fecal egg count reduction test on day zero (0) and day 14 post-treatment by subcutaneously injecting Tectin[®] (each 1 ml contains 10 mg Ivermectin) 0.2 mg/kg B.W. Moreover, the World Association of Advancement for Veterinary Parasitology (WAAVP) guidelines were referenced to detect the resistance. The overall prevalence of gastrointestinal nematodes was 47%, comprising *Haemonchus* spp. 6.66%, *Ostertagia* spp. 8.66%, *Nematodirus* spp. 14%, and *Trichostrongylus* spp. 17.66%. The efficacy of Ivermectin (71%) and the lower limit confidence interval (95%) was 46%, indicating AR to Ivermectin. As documented above, resistance of nematodes to anthelmintic classes, including Ivermectin, could potentially serve as a risk role in propagation, consequence, and maintaining parasitic burden to the productivity of ruminant farms, in addition to solidification set for hindering, monitoring and awareness of the sources of resistance being recommended.

Keywords: *Ivermectin, Sheep, Nematodes, Resistance, Efficacy.*

©Authors, 2025, College of Veterinary Medicine, University of Al-Qadisiyah. This is an open access article under the CC BY 4.0 license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction In equatorial and subtropical territories, gastrointestinal nematode (GIN) have significant health influence in small ruminants, inducing significant production losses (1,2). Clinical and economic risks of these nematodes are usually due to sustained weight, death, decreased food intake, anemia, and retarded weight gain are considerably the most clinical signs (3,4,5). Moreover, diminutive feeding capacity affects absorption of nitrogen, energy, and mortality in heavily infested animals (2).

The ruminal-intestinal nematodes genera described as important parasites in small ruminants (6). Notably, hit with further one species occur commonly, and consequently, complexity of the outcomes and treatment procedures elevated (7).

In human and veterinary clinic, Ivermectin is a confirmed medication for domination parasitic attacks (8). The drug belongs to the avermectins group, which that comprises of 16 macrocyclic lactone

member compounds found in the Institute of Kitasato in Japan in 1967 extracted from *Streptomyces avermitilis*, and thereafter registered in the World Health Organization (9,10,11). Ivermectin is a positive synergistic modulator that eclectically opens inhibitory glutamate-gated chloride ion channels and induces suppression of pharyngeal pumping, motility, and egg release in the reproductive system. (8,12,13,14). Worldwide, it was announced that the extreme and inconsequential use of antiparasitic for abundant periods participated in developing parasite resistant phenomena, which could constrain global food security (15,16). Negatively, documented literatures around the world increased (17,18). Available medication for gastrointestinal nematodes basically relies on repellent drugs, but repeated administration faces multiple resistant parasites. Awareness about anthelmintic resistance and its genetic mechanisms requires more and continual detection for different medications (19).

The fecal egg count reduction test (FECRT) one of the frequent modes utilized to reveal of anthelmintic impedance performed in conjunction with coprocultures to investigate the encompassed parasites (20,21). Recently molecular techniques were used for detection of gastrointestinal nematodes such as *Haemonchus* spp. (22,23).

Small ruminants, including sheep, have a decisive service in promoting the socio-economic revival in abundant countries (24). On account of the productivity regimens, they can be hindered by multiple influences, including infestations of external and internal parasites. So, this work attempted to find out the resistance issue of Ivermectin using the reduction test observed in sheep in Mosul city, Iraq.

Materials and Methods

Ethical approve

This research approved by UM.VET.2024.098

Animal of study

A 730 local male sheep from six private flocks (A, B, C, D, E, and F) raised for fattening purposes participated in this study from various locations in Mosul between October 2024 and February 2025. The animals were \geq seven months, the farm size was between 75 and 150, and there was no history of deworming. The most significant clinical indications, history, and epidemiological data were documented in a clinical card provided by the producers during the sampling procedure.

Fecal samples collection and handling

Three hundred (300) individual fresh fecal samples under septic conditions were obtained and spotted in containers, carried to the laboratory for examination. For detection of ruminal-intestinal nematodes, flotation, sedimentation, and McMaster techniques were used according to Sabatini *et al.* (25). The distinguishing of the nematode eggs done according to Mahmood *et al.* (26).

Fecal egg count and fecal egg count reduction test (FECRT)

Fecal egg counts (FEC) were employed by flotation method protocol (27). McMaster egg counting technique, degree of infestation, and eggs per gram (EPG) were categorized according to Soulsby and Helminths, (28). $EPG = (\text{eggs of 2 pads}) \times 50$ (29,30).

To investigate the efficiency of Ivermectin, through surveying a flock of sheep ($n=75$ and $EPG \geq 200$) was selected. 50 sheep were marked, and further, the selected animals were given Ivermectin (Tectin® 1%) (31). Fecal egg count and EPG were ranked in day Zero (control group T1) prior to injection and on day 14 (treated group T2) for the next medication (32). The equation $FECRT\% = (1 - T2/T1) \times 100$ was utilized (20,21,33).

Statistical analysis

The status of ruminal-intestinal nematodes in sheep was analyzed in Excel sheath software for Windows 7. Significant divergence between T1 and T2 models, FECRT, and lower limit for confidence intervals of 95% were dialed in SPSS software. The definitive decision for the presence of resistance relies on WAAVP directions in which the farm was assessed as resistant if the egg reduction rate was $<95\%$, the lower limit of the 95% confidence interval was $<90\%$. Moreover, the resistance was doubtful when the scores didn't meet (34,35).

Results

Fecal analysis of the sheep samples for the current survey revealed that the overall prevalence of ruminal-intestinal nematodes was 47% (141/300), comprising *Haemonchus* spp. 6.66% (20/300), *Ostertagia* spp. 8.66% (26/300), *Nematodirus* spp. 42/300 (14%), and *Trichostrongylus* spp. 17.66% (53/300) (Table 1) (Figure 1).

Table 1: The status of ruminal-intestinal nematodes in sheep by McMaster method

Type of nematodes	No. of samples examined	No. positive samples	Prevalence %
Haemonchus spp.	300	20	6.66
Ostertagia spp.		26	8.66
Nematodirus spp.		42	14
Trichostrongylus spp.		53	17.66
Overall		141	47

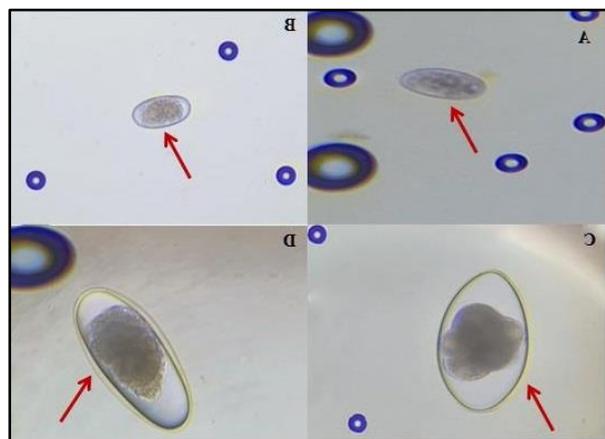


Figure 1: Ruminal-intestinal nematode eggs: A. Haemonchus spp., B. Ostertagia spp., C. Nematodirus spp. D. Trichostrongylus spp. under microscopic examination X400.

Considerably, Ivermectin lowers the fecal eggs on treated animals parallel to the day 0 (Table 2). The outcome also counted a 71% rate of FERCT, and the lower limit of 95% was 46% (Table 2). This finding indicates that ruminal-intestinal nematodes are resistant to Ivermectin.

Table 2: Limit confidence level of 95% in sheep, n=50

Drug	No. animals	± EPG (T1)	± EPG (T2)	FECRT %	CI 95 %
Ivermectin	50	211±201.38 ^a	62± 39.82 ^b	71%	46%

± mean and standard error of mean, CI confidence interval, letter a and b significant difference at P<0.05

Discussion

The gastro-intestinal nematodes have a direct and indirect economic impact on farms productivity worldwide, especially in poorly managed areas, through interference with weight, milk, expense fees for medications, and mortality in heavily infected animals (36-38). In the current study, the overall prevalence of ruminal-intestinal nematodes was 47%. This finding was nearer than the total infection rate reported in the cities of Mosul and Erbil which was 46.49% (39). In the literature, the infection rate of was 42.85% (40), 38% (41), and 30.73% (42). Further, in Sulaymaniyah province was 78.15% (43). There are several works that have reported the infection rate of nematodes in sheep in different countries, such as in Turkey, which was 76% (44), in Iran, which was 94% (45), in China, which was 55.7% (46), and in Pakistan, which was 72.92% (47). The prevalence of nematodes in sheep differs among countries due to different management practices, sample size, efficacy of laboratory tests used, and seasonal climatic variations between countries (48-50). Numerous nematode species were detected in sheep in this study, such as Haemonchus spp., Ostertagia spp., Nematodirus spp., and Trichostrongylus spp. These results agree with Khalaf et al. (39), Moosa et al. (41), and Jwher et al. (42), who reported all these nematode species in Nineveh province, Iraq.

Results showed that the Ivermectin significantly lowered the rate of eggs in the feces in the post-treatment group in contrast with egg excretion to the pre-injection time with reduction rates (71%). These results match the earliest literature vision (31,32). It has been known that Ivermectin has a broad parasitocidal vigor versus round alimentary parasites and skin mange and lice (14,51). Burgess et al., (52) revealed that Ivermectin still the major available antiparasitic preference in the livestock population, mainly for deworming of roundworms and macrocyclic lactone members, as yet is the generally an anthelmintic purpose in the Great British sheep. Cattle farms of the United States of America (53), as well as in Iraq (31). Outcome also points out that the rate of egg sliding is 71%, the lower limit confidence level of 95% was 46%, and based on the WAAVP reference, this view indicates resistance to Ivermectin. Our finding is parallel to previous investigations (6,18,54,55). Presence of anthelmintics resistance could be due to widespread, frequent,

uncontrolled use and incorrect doses of common anthelmintic. It has been clarified that the promiscuous application, readily obtainable, short and frequent period application, re-administration of multiple drug classes under one-year time, and mass treatment (56-58). Moreover, gene expression alternation (59,60). In contrast to the current result, Shihab and Hassan, (31) indicate that the ruminal-intestinal nematodes resistant to Ivermectin were suspected. Generally speaking, mass planning could participate in the development of resistance, which is currently a global issue of concern (61,62).

Conclusions

Current article is premier report consider the resistance feature of Ivermectin in naturally infected sheep with ruminal-intestinal nematodes in Mosul city, Iraq. Ivermectin known to be frequently choice therapy used by veterinarians and farmers against gastrointestinal and ecto-parasites of small and large ruminants in Mosul. Resistance of ruminal-intestinal nematodes to anthelmintic classes, including Ivermectin, could potentially serve as a risk role in consequence and maintaining parasitic burden to the productivity of ruminant farms. In addition, establishing rational measures for prevention, control, and awareness of the sources of resistance is recommended.

Acknowledgment

Grateful thanks to the Mosul College of Veterinary Medicine for prop.

Conflict of Interest

This article's authors are all contented, and there are no conflicts.

Funding source

This research had no specific fund; however, it was self-funded by the authors.

References

1-Besier RB, Kahn LP, Sargison ND, VanWyk JA. Chapter four - the pathophysiology, ecology and epidemiology of *Haemonchus contortus* infection in small ruminants. *Adv Parasitol.* 2016;93: 95-143. DOI:[10.1016/bs.apar.2016.02.022](https://doi.org/10.1016/bs.apar.2016.02.022).

2-Adduci I, Sajovitz F, Hinney B, Lichtmannsperger K, Joachim A, Wittek T, Yan S. Haemonchosis in sheep and goats, control strategies and development of vaccines against *Haemonchus contortus*. *Animals.* 2022;12(18):2339-23358. DOI:[10.3390/ani12182339](https://doi.org/10.3390/ani12182339) PMID: 36139199.

3-Hoberg EP, Zarlenga DS. Evolution and Biogeography of *Haemonchus contortus*: Linking Faunal Dynamics in Space and Time. *Adv Parasitol.* 2016, 93, 1–30. [Google Scholar]

4-Carvalho N, Neves JH, Pennacchi CS, Castilhos AM, Amarante AFT. Performance of lambs under four levels of dietary supplementation and artificially mix-infected with *Haemonchus contortus* and *Trichostrongylus colubriformis*. *Rev Bras Parasitol Vet.* 2021; 30(1): e025420. DOI: [10.1590/S1984-29612021010](https://doi.org/10.1590/S1984-29612021010).

5-Höglund J, Baltrušis P, Enweji N, Gustafsson K. Signs of multiple anthelmintic resistance in sheep gastrointestinal nematodes in Sweden. 2022;36: 1-9. DOI:[10.1016/j.vprsr.2022.100789](https://doi.org/10.1016/j.vprsr.2022.100789).

6-Hassan NMF, Ghazy AA. Advances in diagnosis and control of anthelmintic resistant gastrointestinal helminths infecting ruminants. *J Parasit Dis.* 2022;46(3):901-915. DOI:[10.1007/s12639-021-01457-z](https://doi.org/10.1007/s12639-021-01457-z).

7-Roeber F, Jex AR, Gasser RB. Advances in the diagnosis of key gastrointestinal nematode infections of livestock, with an emphasis on small ruminants. *Biotechnol Adv.* 2013;31(8): 1135-1152. DOI: [10.1016/j.biotechadv.2013.01.008](https://doi.org/10.1016/j.biotechadv.2013.01.008)

8-Laing R, Gillan V, Devaney E. Ivermectin -Old Drug, New Tricks?. *Trends Parasitol.* 2017; 33(6): 463-472. DOI:[10.1016/j.pt.2017.02.004](https://doi.org/10.1016/j.pt.2017.02.004)

9-Canga GA, Sahagún Prieto AM, Díez Liébana MJ, Martínez NF, Vega MS, García Vieitez JJ. The pharmacokinetics and interactions of ivermectin in humans—a mini-review. *AAPS J.* 2008;10(1):42-46. <https://doi.org/10.1208/s12248-007-9000-9>.

10-Crump A, Ōmura S. Ivermectin, 'wonder drug' from Japan: the human use perspective. *Proc Jpn Acad Ser B Phys Biol Sci.* 2011;87(2):13-28. DOI: [10.2183/pjab.87.13](https://doi.org/10.2183/pjab.87.13).

11-Kircik LH, Del Rosso JQ, Layton AM, Schaubert J. Over 25 years of clinical experience with ivermectin: an overview of safety for an increasing number of indications. *J Drugs Dermatol.* 2016;15:325-332. <https://pubmed.ncbi.nlm.nih.gov/26954318/>.

12-Li BW, Rush AC, Weil GJ. High level expression of a glutamate-gated chloride channel gene in reproductive tissues of *Brugia malayi* may explain the sterilizing effect of ivermectin on filarial worms. *Int J Parasitol Drugs Drug Resist.* 2014; 4(2):71-76. DOI: [10.1016/j.ijpddr.2014.01.002](https://doi.org/10.1016/j.ijpddr.2014.01.002).

13-Ballesteros C. 1, Lucienne Tritten 1, Maeghan O'Neill 1, Erica Burkman 2 3, Weam I Zaky 3 4, Jianguo



- Xia 1, Andrew Moorhead 2 3, Steven A Williams 3, Timothy G Gear. The Effects of Ivermectin on *Brugia malayi* Females In Vitro: A Transcriptomic Approach. *PLoS Negl Trop Dis.* 2016 16;10(8):e0004929. DOI: [10.1371/journal.pntd.0004929](https://doi.org/10.1371/journal.pntd.0004929).
- 14-Martin RJ, Robertson AP, Choudhary S. Ivermectin: An Anthelmintic, an Insecticide, and Much More. *Trends Parasitol.* 2021; 37(1): 48-64. DOI:[10.1016/j.pt.2020.10.005](https://doi.org/10.1016/j.pt.2020.10.005).
- 15-Charlier J, Morgan ER, Rinaldi L, van Dijk J, Demeler J, Höglund J, Hertzberg H, Van Ranst B, Hendrickx G, Vercruysse J, Kenyon F. Practices to optimise gastrointestinal nematode control on sheep, goat and cattle farms in Europe using targeted (selective) treatments. *Vet Rec.* 2014; 175(10):250-255. DOI: [10.1136/vr.102512](https://doi.org/10.1136/vr.102512).
- 16-Rose Vineer H, Morgan ER, Hertzberg H, Bartley DJ, Bosco A, Charlier J, Chartier C, Claerebout E, de Waal T, Hendrickx G, Hinney B, Höglund J, Jezek J, Kašný M, Keane OM, Martínez-Valladares M, Mateus TL, McIntyre J, Mickiewicz M, Munoz AM, Phythian CJ, Ploeger HW, Rataj AV, Skuce PJ, Simin S, Sotiraki S, Spinu M, Stuen S, Thamsborg SM, Vadlejch J, Varady M, von Samson-Himmelstjerna G, Rinaldi L. Increasing importance of anthelmintic resistance in European livestock: creation and meta-analysis of an open database. *Parasite.* 2020;27: 69-83. DOI:[10.1051/parasite/2020062](https://doi.org/10.1051/parasite/2020062).
- 17-Gilleard JS, Kotze AC, Leathwick D, Nisbet AJ, McNeilly TN, Besier B. A journey through 50 years of research relevant to the control of gastrointestinal nematodes in ruminant livestock and thoughts on future directions. *Int J Parasitol.* 2021;51(13):1133-1151. DOI: [10.1016/j.ijpara.2021.10.007](https://doi.org/10.1016/j.ijpara.2021.10.007)
- 18-Bassetto CC, Albuquerque ACA, Lins JGG, Marinho-Silva NM, Chocobar MLE, Bello HJS, Mena MO, Niciura SCM, Amarante AFT, Chagas ACS. Revisiting anthelmintic resistance in sheep flocks from São Paulo State, Brazil. *Int J Parasitol: Drugs Drug Resist.* 2024; 24:100527, DOI:[10.1016/j.ijpddr.2024.100527](https://doi.org/10.1016/j.ijpddr.2024.100527)
- 19-Kotze AC, Hunt PW, Skuce P, Samson-Himmelstjerna G, Martin RJ, Sager H, Krućken J, Hodgkinson J, Lespine A, Jex AR, Gilleard JS, Beech RN, Wolstenholme AJ, Demeler J, Robertson AP, Charvet CL, Neveu C, Kaminsky R, Rufener L, Alberich M, Menez C, Prichard RK. Recent advances in candidate genes and whole-genome approaches to the discovery of anthelmintic resistance markers and the description of drug/receptor interactions. *Int J Parasitol Drugs Drug Resist.* 2014; 4(3):164-184. DOI: [10.1016/j.ijpddr.2014.07.007](https://doi.org/10.1016/j.ijpddr.2014.07.007).
- 20-Coles G, Jackson F, Pomroy W, Prichard R, von Samson-Himmelstjerna G, Silvestre A, Taylor M, Vercruysse J. The detection of anthelmintic resistance in nematodes of veterinary importance. *Vet Parasitol.* 2006;136(3-4):167-185. DOI: [10.1016/j.vetpar.2005.11.019](https://doi.org/10.1016/j.vetpar.2005.11.019).
- 21-Levecke B, Speybroeck N, Dobson RJ, Vercruysse J, Charlier J. Novel Insights in the Fecal Egg Count Reduction Test for Monitoring Drug Efficacy against Soil-Transmitted Helminths in Large-Scale Treatment Programs. *PLoS Negl Trop Dis.* 2011;5(12): e1427. DOI:[10.1371/journal.pntd.0001427](https://doi.org/10.1371/journal.pntd.0001427)
- 22-Abed AN, Alfatlawi MAA. *Haemonchus contortus*: Conventional and molecular identification and characterization via sequencing-based phylogenetic analysis in sheep. *Al-Qadisiyah J Vet Med Sci.* 2025; 24 (1): 20-25. DOI:[10.29079/qjvms.2024.151711.1029](https://doi.org/10.29079/qjvms.2024.151711.1029)
- 23-Mohammed NQ, Ahmed HS, Jarad NI, Al-Shabbani AH. *Haemonchus contortus*: Review of recent molecular advances in anthelmintic resistance and vaccination. *Al-Qadisiyah J Vet Med Sci.* 2020;19(2):15-26. DOI: [10.29079/qjvms.2020.179345](https://doi.org/10.29079/qjvms.2020.179345)
- 24-Keagan JD, Keane OM, Good B, DeWaal T, Denny M, Hanrahan JP, Fitzgerald W, Sheehan M. A nationwide survey of anthelmintic treatment failure on sheep farms in Ireland. *Irish Vet J.* 2017;70(7):1-8. DOI: [10.1186/s13620-017-0086-9](https://doi.org/10.1186/s13620-017-0086-9).
- 25-Sabatini GA, de Almeida Borges F, Claerebout E, Gianechini LS, Höglund J, Kaplan RM, Zanetti-Lopes WD, Mitchell S, Rinaldi L, von Samson-Himmelstjerna G, Stefan P, Woodgate R. Practical guide to the diagnostics of ruminant gastrointestinal nematodes, liver fluke and lungworm infection: interpretation and usability of results. *Parasit Vectors.* 2023;16(1): 58-47. DOI:[10.1186/s13071-023-05680-w](https://doi.org/10.1186/s13071-023-05680-w).
- 26-Mahmood O, Muhsin S, Hussein M. Morphological Diagnosis for Some Eggs of Gastrointestinal Nematodes from Sheep. *Tikrit j eng sci.* 2019;19(3): 6-9. DOI:[10.25130/tjas.19.3.2](https://doi.org/10.25130/tjas.19.3.2)
- 27-Tomczuk K, Kostro K, Szczepaniak KO, Grzybek M, Studzińska M, Demkowska-Kutrzepa M, Roczeń-Karczmarz M. Comparison of the sensitivity of coprological methods in detecting Anoplocephala perfoliata invasions. *Parasitol Res.* 2014;113(6): 2401-2406. DOI: [10.1007/s00436-014-3919-4](https://doi.org/10.1007/s00436-014-3919-4)
- 28-Soulsby EJW. *Helminths, Arthropods and Protozoa of Domesticated Animals*, Seventh Edition. Bailliere Tindall, London: Lea and Febiger, Philadelphia. 1982;



212-258 P. <https://academic.oup.com/trstmh/article-abstract/78/3/329/1888909>.

29-Hendrix CM. Diagnostic veterinary parasitology. 2nd ed. USA. 1998; 240-244 P.

30-Pinilla León JC, Delgado NU, Florez AA. Prevalence of gastrointestinal parasites in cattle and sheep in three municipalities in the Colombian Northeastern Mountain, *Veterinary World*, 2019;12(1): 48-54. DOI: [10.14202/vetworld.2019.48-54](https://doi.org/10.14202/vetworld.2019.48-54)

31-Shihab HH, Hassan SD. Detection of resistance against anti-helminths drugs in gastrointestinal nematodes of calves using fecal egg count reduction test FECRT., *Iraqi J Vet Sci*. 2023; 37(1): 283-288. DOI: [10.33899/ijvs.2022.134037.2333](https://doi.org/10.33899/ijvs.2022.134037.2333)

32-Mushonga AN, Washaya S, Nyamushamba GB. Resistance of Gastrointestinal Nematodes to Anthelmintics in Sheep Production in Zimbabwe. *Farm Anim Health Nutr*. 2024; 3(1): 22-27. DOI: [10.58803/fahn.v3i1.39](https://doi.org/10.58803/fahn.v3i1.39)

33-Wondimu A, Bayu Y. Anthelmintic Drug Resistance of Gastrointestinal Nematodes of Naturally Infected Goats in Haramaya, Ethiopia. *J Parasitol Res*. 2022; 4025902:1-7. DOI: [10.1155/2022/4025902](https://doi.org/10.1155/2022/4025902)

34-Coles GC, Bauer C, Borgsteede FH, Geerts S, Klei TR, Taylor MA, Waller PJ. World Association for the Advancement of Veterinary Parasitology (WAAVP) methods for the detection of anthelmintic resistance. *Vet Parasitol*. 1992; 44(1-2):35-44. DOI: [10.1016/0304-4017\(92\)90141-u](https://doi.org/10.1016/0304-4017(92)90141-u).

35-Herrera-Manzanilla FA, Ojeda-Robertos NF, González-Garduño R, Cámara-Sarmiento R, Torres-Acosta JFJ. Gastrointestinal nematode populations with multiple anthelmintic resistance in sheep farms from the hot humid tropics of Mexico. *Vet Parasitol Reg Stud Rep*. 2017;9:29-33. DOI: [10.1016/j.vprsr.2017.04.007](https://doi.org/10.1016/j.vprsr.2017.04.007)

36-Martínez-Valladares M, Geurden T, Bartram DJ, Martínez-Pérez JM, Robles-Pérez D, Bohórquez A, Florez E, Meana A, Rojo-Vázquez F. Resistance of gastrointestinal nematodes to the most commonly used anthelmintics in sheep, cattle and horses in Spain. *Vet Parasitol*. 2015; 211: 228-233. DOI: [10.1016/j.vetpar.2015.05.024](https://doi.org/10.1016/j.vetpar.2015.05.024)

37-Charlier J, Rinaldi L, Musella V, Ploeger HW, Chartier C, Vineer HR, Hinney B, von Samson-Himmelstjerna G, Băcescu B, Mickiewicz M, Mateus TL, Martínez-Valladares M, Quealy S, Azaizeh H, Sekovska B, Akkari H, Petkevicius S, Hektoen L, Höglund J, Morgan ER, Bartley DJ, Claerebout E. Initial assessment of the economic burden of major parasitic

helminth infections to the ruminantlivestock industry in Europe. *Prev Vet Med*. 2020; 182:105103. DOI: [10.1016/j.prevetmed.2020](https://doi.org/10.1016/j.prevetmed.2020).

38-Charlier J, Williams DJ, Ravinet N, Claerebout E. To treat or not to treat: Diagnostic thresholds in subclinical helminth infections of cattle. *Trends Parasitol*. 2022; 39: 139-151. DOI: [10.1016/j.pt.2022.11.014](https://doi.org/10.1016/j.pt.2022.11.014)

39-Khalaf WK, Jarjees MT, Hasan MH. Prevalence of Gastrointestinal Nematode of Sheep in Mosul and Erbil City. *Al-Anbar J Vet Sci*. 2023;16(2):44-53. DOI: [10.37940/AJVS.2023.16.2.5](https://doi.org/10.37940/AJVS.2023.16.2.5)

40-Sulaiman EG, Talib Q, Daham E, Arsalan SH. Study of some eggs and oocysts of internal parasites in sheep in Mosul. *Iraqi J Vet Sci*. 2005;19:21-32. DOI: [10.33899/ijvs.2005.37275](https://doi.org/10.33899/ijvs.2005.37275)

41-Moosa DA, Hussien AM, Hameed HM, Hasan SA. Diagnostic and hematological study in sheep infected with gastrointestinal nematode in Mosul City. *Al-Anbar J Vet Sci*. 2022;15(1):29-33. DOI: [10.37940/AJVS.2022.15.1.4](https://doi.org/10.37940/AJVS.2022.15.1.4).

42-Jwher DhM, Jarjees MT, Shareef AM. A study of the gastrointestinal parasites in Awssi sheep and surrounding environment. *Iraqi J Vet Sci*. 2021; 35(3): 561-567. DOI: [10.33899/ijvs.2020.127174.1478](https://doi.org/10.33899/ijvs.2020.127174.1478)

43-Mohammed AA. Prevalence of haemoprotozoan and gastrointestinal parasites of sheep imported from Syria into Sulaymaniyah province of Iraq. *Ann Parasitol*. 2021; 67(3): 465-471. DOI: [10.17420/ap6703.359](https://doi.org/10.17420/ap6703.359)

44-Altaş M, Sevgili M, Gökçen A, Bayburs, HC. Prevalence of gastrointestinal nematodes in sheep in the Sanliurfa Region. *Turkiye Parazitol Derg*. 2006;30(4):317-321. <https://pubmed.ncbi.nlm.nih.gov/17309037/>

45-Hatam-Nahavandi K, Carmena D, Rezaeian M, Mirjalali H, Rahimi HM, Badri M, Vafae Eslahi A, Shahrivar FF, Oliveira SRM, de Lourdes Pereira M, Ahmadpour E. Gastrointestinal parasites of domestic mammalian hosts in Southeastern Iran. *Vet Sci*. 2023; 10(4): 261-275. DOI: [10.3390/vetsci10040261](https://doi.org/10.3390/vetsci10040261)

46-Cai W, Cheng C, Feng Q, M, Y, Hua E, Jiang S, Hou Z, Liu D, Yang A, Cheng D, Xu J, Tao J. Prevalence and risk factors associated with gastrointestinal parasites in goats (*Capra hircus*) and sheep (*Ovis aries*) from three provinces of China. *Front Microbiol*. 2023;14:1287835. DOI: [10.3389/fmicb.2023.1287835](https://doi.org/10.3389/fmicb.2023.1287835).

47-Rizwan HM, Zohaib HM, Sajid MS, Tahir UB, Kausar R, Nazish N, Ben Said M, Anwar N, Maqbool M, Fouad D, Ataya FS. Unveiling the hidden threat: investigating



- gastrointestinal parasites and their costly impact on slaughtered livestock. *Rev Bras Parasitol Vet.* 2024; 33(3): e007224. DOI: [10.1590/S1984-2961202406148](https://doi.org/10.1590/S1984-2961202406148)-Hunde FT, Chali AR. Study on prevalence of major gastrointestinal nematodes of sheep in Wayu Tuka and Diga District, Oromia Regional State. *Vet Med Open J.* 2021; 6(1): 13-21. DOI: [10.17140/VMOJ-6-15449](https://doi.org/10.17140/VMOJ-6-15449)-Desalegn C, Berhanu G. Assessment of the epidemiology of the gastrointestinal tract nematode parasites in sheep in Toke Kutaye, West Shoa Zone, Ethiopia. *Vet Med.* 2023; 14:177-183. DOI: [10.2147/VMRR.S427828](https://doi.org/10.2147/VMRR.S427828)
- 50-Solomon L, Haile G, Ahmed NA, Abdeta D, Galalcha W, Hailu Y. Epidemiology and field efficacy of anthelmintic drugs associated with gastrointestinal nematodes of sheep in Nejo district, Oromia, Ethiopia. *Sci Rep.* 2024; 14(1): 6841. DOI: [10.1038/s41598-024-55611-7](https://doi.org/10.1038/s41598-024-55611-7)
- 51-Bauck S. Ivermectin toxicity in small animals. *Can Vet J.* 1987;28(9): 563-564. <https://pubmed.ncbi.nlm.nih.gov/articles/PMC1680532/>
- 52-Burgess CG, Bartley Y, Redman E, Skuce PJ, Nath M, Whitelaw F, Tait A, Gilleard JS, Jackson F. A survey of the trichostrongylid nematode species present on UK sheep farms and associated anthelmintic control practices. *Vet Parasitol.* 2012;189(2-4):299-307. DOI: [10.1016/j.vetpar.2012.04.009](https://doi.org/10.1016/j.vetpar.2012.04.009).
- 53-McArthur MJ, Reinemeyer CR. Herding the US cattle industry toward a paradigm shift in parasite control. *Vet Parasitol.* 2014;204(1-2):34-43. DOI: [10.1016/j.vetpar.2013.12.021](https://doi.org/10.1016/j.vetpar.2013.12.021)
- 54-Kumar S, Singh S. Detection Of Multiple Anthelmintic Resistance Against Gastrointestinal Nematodes In Sheep On Central Sheep Breeding Farm, Hisar. *Haryana Vet.* 2016; 55 (2):210-213. <https://www.cabidigitallibrary.org/doi/pdf/10.5555/20173200084>
- 55-Mondragón-Ancelmo J, Olmedo-Juárez A, Reyes-Guerrero DE, Ramírez-Vargas G, Ariza-Román AE, López-Arellano ME, de Gives PM, Napolitano F. Detection of Gastrointestinal Nematode Populations Resistant to Albendazole and Ivermectin in Sheep. *Animals.* 2019; 9(10): 775-784. DOI:[10.3390/ani9100775](https://doi.org/10.3390/ani9100775)
- 56-Sargison ND, Jackson F, Bartley DJ, Wilson DJ, Stenhouse LJ, Penny CD. Observations on the emergence of multiple anthelmintic resistance in sheep flocks in the South-East of Scotland. *Vet Parasitol.* 2007; 145: 65-76. DOI: [10.1016/j.vetpar.2006.10.024](https://doi.org/10.1016/j.vetpar.2006.10.024)
- 57-Silva FF, Bezerra HMFF, Feitosa TF, Vilela VLR. Nematode resistance to five anthelmintic classes in naturally infected sheep herds in Northeastern Brazil. *Rev Bras Parasitol Vet.* 2018;27(4): 423-429. DOI:[10.1590/S1984-296120180071](https://doi.org/10.1590/S1984-296120180071)
- 58-Billingsley P, Binka F, Chaccour C, Foy BD, Gold S, Gonzalez-Silva M, Jacobson J, Jagoe G, Jones C, Kachur P, Kobylinski K, Last A, Lavery JV, Mabey D, Mboera D, Mbogo C, Mendez-Lopez A, Rabinovich NR, Rees S, Richards F, Rist C, Rockwood J, Ruiz-Castillo P, Sattabongkot J, Saute F, Slater H, Steer A, Xia K, Zullinger R. A Roadmap for the development of Ivermectin as a complementary malaria vector control tool. *Am J Trop Med Hyg.* 2022; 102(2s):3-24. DOI: [10.4269/ajtmh.19-0620](https://doi.org/10.4269/ajtmh.19-0620)
- 59-Khangembam R, Singh H, Jyoti RSS, Singh NK. Effect of synergists on ivermectin resistance in field populations of *Rhipicephalus (Boophilus) microplus* from Punjab districts, India. *Ticks Tick Borne Dis.* 2018; 9(3):682-686. DOI: [10.1016/j.ttbdis.2018.02.005](https://doi.org/10.1016/j.ttbdis.2018.02.005)
- 60-Kim JH, Gellatly KJ, Lueke B, Kohler M, Nauen R, Murenzi E, Yoon KS, Clark JM. Detoxification of ivermectin by ATP binding cassette transporter C4 and cytochrome P450 monooxygenase 6CJ1 in the human body louse. *Pediculus Humanus Humanus Insect Mol Biol.* 2018;27(1):73-82. DOI: [10.1111/imb.12348](https://doi.org/10.1111/imb.12348)
- 61-Osei-Atweneboana MY, Awadzi K, Attah SK, Boakye DA, Gyapong JO, Prichard1 RK. Phenotypic Evidence of Emerging Ivermectin Resistance in *Onchocerca volvulus*. *PLOS Negl Trop Dis.* 2011; 5(3): e998. DOI:[10.1371/journal.pntd.0000998](https://doi.org/10.1371/journal.pntd.0000998)
- 62-Nana-Djeunga HC, Bourguinat C, Pion SD, Bopda J, Kengne-Ouafo JA, Njiokou F, Prichard RK, Wanji S, Kamgno J, Boussinesq M. Reproductive status of *Onchocerca volvulus* after ivermectin treatment in an ivermectin-naive and a frequently treated population from Cameroon. *PLoS Negl Trop Dis.* 2014;24;8(4):e2824. DOI: [10.1371/journal.pntd.0002824](https://doi.org/10.1371/journal.pntd.0002824)