



Review article:

Mycoplasma gallisepticum Infection in Poultry: Article Review

Ansam Khalid Mohammed

**Department of Microbiology-College of Veterinary Medicine-University of Baghdad-
Baghdad, Iraq.**

ORCID: 0000-0002-6374-3253

E-mail: ansam@covm.uobaghdad.edu.iq

Abstract

Mycoplasma gallisepticum (MG) is the most important causative factor of respiratory infections in chickens and turkey resulting chronic respiratory disease CRD and Infectious sinusitis respectively. These diseases transmitted throughout the world in different ages of to all nearby bird species. The infected poultry become carrier that can infect all birds around world. This review introduces a short information about MG infection specially in avian species. The detection of this causative agent made by isolation of MG by serological test or culturing with media containing digested protein, meat infusion base with serum, yeast factors, glucose and bacterial inhibitors on many commercial liquid media and agar media used for isolation like: Frey medium and SP-4 medium and other isolation methods like rapid agglutination test which made by several methods can be used to diagnosis the MG like: Haemagglutination inhibition, ELISA, PCR and molecular methods to differentiate between MG strains, DNA fingerprinting that uses RAPD (random amplified polymorphic DNA), GTS (gene targeting sequencing) for typing strains.

Keywords: *Mycoplasma gallisepticum*; Avian; Respiratory diseases.

Introduction

With a world-wide distribution, *Mycoplasma gallisepticum* (MG) has the greatest significant infections that cause high economic losses in avian species, as highly transmissible avian pathogen (1). It is the primary causes of chronic respiratory diseases (CRD) infection in chicks and infectious sinusitis in turkey, of all types and ages. *M. gallisepticum* infection is spread horizontally from one fowl to another; moreover, it can be vertically (1). Primary description of the disease by Dodd 1905 (2) in England named “epizootic pneumoenteritis” was observed in turkeys (2). *M. gallisepticum* diseases are specifying by respiration rales, nasal and ocular discharge, coughing and conjunctivitis. In the turkey's species, often infraorbital sinusitis is present (1). Clinical manifestation typically emerges gradually, and the infection may last for a long period. As a result, the carrier bird continues to harbor the disease throughout its lifetime, posing a threat to other

avian populations (3). Decrease in growth and production are prominent in MG infection (4). Initial damage usually be made of heterophils and macrophages, and subsequent damage demonstrate huge quantity of lymphocytes along excessive parts of T-lymphocytes (5). Like the other members of the class Mollicutes, family Mycoplasmataceae, the genus of *Mycoplasma* lacks a cell wall and are smallest self-replicating prokaryotes with reduced genome, with the minimal genetic information, many metabolic pathways and the capability of growing on artificial cell-free media (6). These characters relate widely to MG and the host species, moreover, the organism is very sensitive to nature where there is difficult in culturing, selective antibiotic sensitivity, inhibition of phagocytosis, and its intimate association with host cells. MG constitutes of two duplicate of a 23S-5S rRNA gene aggregation, both not found close to single 16S rRNA gene (7). *M.*



gallisepticum was first identified as serotype A (8 & 9) to distinguish it from other avian mycoplasmas (10). By 1993, molecular techniques helped distinguish between mycoplasmas with antigenic and physical similarities (11). *M. gallisepticum* infections may be complicated by other respiratory virus infections such as infectious bronchitis (IB), Newcastle disease virus (NDV), Avian Influenza virus (AIV) and very often along with a secondary infection of *Escherichia coli*; Haemophilus, or avian rhinotracheitis virus causes severe air-sacculitis, also known as “Air sac disease”, leading to aggravated clinical CRD, high morbidity, mortality and/or increased condemnations at processing (12). *Mycoplasma gallisepticum* features a small genomic size comprised of only 996442 bp nucleotides for Rlow strain (13) with limited biosynthetic capabilities. MG membrane proteins are crucial in establishing MG morphology, movement and colonization of the host (14 & 15). As an opportunistic pathogen, MG depends on its parasitic lifestyle and despite several limitations, their degenerative evolution allow them to transmitted through immobile surfaces, such as glass, plastic and eukaryotic cells, absence of motion adjacent such as flagella or pili (16). Antigenic variation, phase variation, superantigens are a few of the mechanisms adopted by MG to evade the host’s lymphatic system (1). In chickens, *M. gallisepticum* is outlined by excessive infection in trachea, air sacs and lungs; conjunctivitis; rales; nasal and mucosal discharge. MG attached to the tracheal epithelial cells mediating filtering of macrophages, heterophils and lymphocytes to the tracheal connective tissue. Yet, molecular mechanism of infectious associated with excessive infectious MG, associated with excessive infection reaction, is elusive (17). Of an infinitesimal size and minimal genetic information, MG lack bacterial cell wall; and hence it is unaffected by β -lactam antimicrobials which target cell wall synthesis. *Mycoplasma gallisepticum* have the ability to penetrate cells; possess a trilaminar membrane and they are highly polymorphic.

MG is facultative anaerobes with optimal temperature of 35-37 C°; and requires an enriched media of 10-20% animal serum and yeast extract. MG causes significant economic losses despite the absence of clinical signs. Condemnations in body, feeding decrease efficiency with decrease of egg producing, reduced hatchability and growth, aggravating and co-existing with other disease agents and increase in medication or vaccinations (11). MG outbreak persists in many countries around the globe and various measures have been enforced with outcomes demonstrated far from satisfactory. Extensive biosecurity and surveillance is a well control MG-program, in the United States, an extensive National Poultry Improvement Plan has been adopted by hatcheries and poultry breeders with success in increasing MG-free flock. The isolation of MG can be done by serological test by detecting DNA directly in infected birds. Culture media contain digested protein, meat infusion base with serum, yeast factors, glucose and bacterial inhibitors (18). Rapid agglutination test one of serological tests that used commonly within three days of sample collection (19). Several methods can be used to diagnosis the MG like: Haemagglutination inhibition, ELISA, PCR and molecular methods can differentiate the MG strains, DNA fingerprinting that uses RAPD (random amplified polymorphic DNA), GTS (gene targeting sequencing) for typing strains (20). Isolation of causative agent was slow requiring three weeks (21), with modification of media was added (22 & 23). Many commercial liquid media and agar media used for isolation like: Frey medium and SP-4 medium, the trachea or choanal cleft or even oviduct swabs can be taken (24). Best growth of MG achieved with 37 C° in high humidity in 5% CO₂, the colonies can be seen after three to five days of incubation (21). The MG classic colonies were small, circular with elevated center like fried egg appearance. Using 16S rRNA gene and sequencing methods with highly specific results, also identification can be done using direct or indirect immune fluorescence techniques,



immune-peroxidase, immunobinding assay. The controls of MG in many countries based on maintain the breeding flocks free from MG infection, the recovered birds had immunity but it still contains the organism. The using of antibiotics was very important in order to prevent attachment of organism with epithelial cells of birds (25). Infection affected cell immune system inducing lymphocytes stimulation (B and T) that lead to produce cytokines (26), natural killer and cytotoxic T cell (27), these inflammatory events occur after interaction of MG with mucosal surface of birds (28). Infiltration of heterophils, macrophages, lymphocytes, polymorphonuclear leukocytes and natural killer also involve in innate and adaptive responses (29). Antibodies in respiratory secretion protect against MG infection by inhibiting the attachment of organism with tracheal epithelia cells (30), by release antibodies including IgA, IgM, IgD, IgE and IgG (31). Vaccinations methods was the best for controlling and maintain the breeding flocks free, there were two types of live vaccines: intranasal or in eye drop and the second type was the fine spray one. Large range of choice were available including vaccination activated, oil emulsion bacterins, live or recombinant live vaccines (32). MG shows sensitivity inside and outside body for many antibiotics like macrolides (erythromycin, tylosin, spiramycin,

licomycine and kitasamycin), tetracyclines (oxytetracycline, chlortetracycline and doxycycline) and fluoroquinolones (imequil, norfloxacin, enrofloxacin and danofloxacin) and tiamulin, the treating birds with antimicrobials decrease the clinical symptoms and the risk of transmission between flocks. Medical plants such as tannins, terpenoids, alkaloids and flavonoids provides good source of anti-infection against microbial agents, and these plants had many compounds treating or protects birds against infection (33). Several studies were done inside Iraq without referring to this important disease (34, 35, 36, 37, 38 & 39). Whereas some of authors published several works deal with this infection in birds (40, 41, 42 & 43).

Conclusion

The MG is the most important causative factor of respiratory infections in chickens and turkey this causative agents could be diagnosed using isolation procedures and molecular or serological methods and can be treated with several medical antibiotics and medical plants and could be controled using vaccination protocol.

Conflict of Interest: The author declares that there is no conflict of interest

References

1. Ramadan NM. Mycoplasma gallisepticum overview in poultry. *Am J Biomed Sci Res.* 2018;4(5):354-355.<https://doi.org/10.34297/AJBSR.2019.04.000833>
2. Dodd S. Epizootic pneumo-enteritis of the turkey. *J Comp Path Therap.* 1905;18:239-245.[https://doi.org/10.1016/S0368-1742\(05\)80041-4](https://doi.org/10.1016/S0368-1742(05)80041-4)
3. Chen H, Yu S, Hu M, Han X, Chen D, Qiu X, Ding C. Identification of biofilm formation by *Mycoplasma gallisepticum*. *Vet Microbiol.* 2012 Jul 17;161(1-2):96-103.<https://doi.org/10.1016/j.vetmic.2012.07.013>
4. Bradbury JM. Poultry mycoplasmas: sophisticated pathogens in simple guise. *Br Poult Sci.* 2005;46:25-136.<https://doi.org/10.1080/00071660500066282>
5. Szczepanek SM, Silbart LK. Host immune responses to mycoplasmas. In: Browning GF, Citti C, editors. *Mollicutes Molecular Biology and Pathogenesis.* Caister Academic Press; 2014. p. 273-288.
6. Semashko T, Arzamasov A, Fisunov GY, Govorun VM. Transcription profiling data set of different states of *Mycoplasma gallisepticum*. *Genomics Data.* 2017;11:49-54.<https://doi.org/10.1016/j.gdata.2016.11.021>
7. Chen X, Finch LR. Novel arrangement of rRNA genes in *Mycoplasma gallisepticum*: separation of the 16S gene of one set from the 23S and 5S genes. *J Bacteriol.* 1989 May;171(5):2876-2878.<https://doi.org/10.1128/jb.171.5.2876-2878.1989>
8. Adler HE, Yamamoto R, Berg J. Strain differences of pleuropneumonia-like organisms of avian origin.



- Avian Dis. 1957;1(1):19-27.<https://doi.org/10.2307/1587542>
9. Yamamoto Y, Adler H. Characteristics of pleuropneumonia-like organisms of avian origin. II. Cultural, biochemical, morphological and further serological studies. *J Infect Dis.* 1958;102(3):243-250.<https://doi.org/10.1093/infdis/102.3.243>
 10. Yoder HW. Characterization of avian *Mycoplasma*. *Avian Dis.* 1964;8:481-512.<https://doi.org/10.2307/1587937>
 11. Bradbury JM, Abdul Wahab OM, Yavari CA, Dupiellet JP, Bové JM. *Mycoplasma imitans* sp. nov. is related to *Mycoplasma gallisepticum* and found in birds. *Int J Sys Bacteriol.* 1993 Oct;43(4):721-728.<https://doi.org/10.1099/00207713-43-4-721>
 12. Sid H, Hartmann S, Petersen H, Ryll M, Rautenschlein S. *Mycoplasma gallisepticum* modifies the pathogenesis of influenza A virus in the avian tracheal epithelium. *Int J Med Microbiol.* 2016;306(3):174-186.<https://doi.org/10.1016/j.ijmm.2016.04.001>
 13. Tan L, Hu M, Yu S, Wang X, Lu F, Qiu X, Song C, Sun Y, Ding C. Characterization of the chaperonin GroEL in *Mycoplasma gallisepticum*. *Arch Microbiol.* 2015 Feb;197(2):235-244.<https://doi.org/10.1007/s00203-014-1047-2>
 14. Jarrell KF, McBride MJ. The surprisingly diverse ways that prokaryotes move. *Nat Rev Microbiol.* 2008 Jun;6(6):466-476.<https://doi.org/10.1038/nrmicro1900>
 15. Panicker IS, Kanci A, Chiu CJ, Veith PD, Glew MD, Browning GF, Markham PF. A novel transposon construct expressing PhoA with potential for studying protein expression and translocation in *Mycoplasma gallisepticum*. *BMC Microbiol.* 2012;12:138.<https://doi.org/10.1186/1471-2180-12-138>
 16. Razin S, Yogev D, Naot Y. Molecular biology and pathogenicity of mycoplasmas. *Microbiol Mol Biol Rev.* 1998 Dec;62(4):1094-1156.<https://doi.org/10.1128/MMBR.62.4.1094-1156.1998>
 17. Majumder S. Role of *Mycoplasma gallisepticum* and Host Airway Epithelial Cell Interaction in Inflammation. [dissertation]. Storrs: University of Connecticut; 2014.
 18. Almanama MA. Prevalence of *Mycoplasma gallisepticum* in the Ten licensed Hatcheries in Gaza strip, Palestine. [thesis]. [Online]. Available from:<https://www.yumpu.com/en/document/view/28809525/prevalence-of-mycoplasma-gallisepticum-in-the-ten-licensed->
 19. Crupper C, Applegate R, Robel R. Prevalence of mycoplasma Antibodies in Lesser Prairie-Chicken Sera. *Avian Dis J.* 2001;3:708-712.[https://doi.org/10.1637/0005-2086\(2002\)046\[0708:POMAIL\]2.0.CO;2](https://doi.org/10.1637/0005-2086(2002)046[0708:POMAIL]2.0.CO;2)
 20. Ferguson N, Hepp D, Sun S, Ikuta N, Levisohn S, Kleven S, García M. Use of molecular diversity of *Mycoplasma gallisepticum* by gene-targeted sequencing and random amplified polymorphic DNA analysis for epidemiological studies. *Microbiol J.* 2005;151:1883-1893.<https://doi.org/10.1099/mic.0.27642-0>
 21. Kleven SH. Control of avian mycoplasma infections in commercial poultry. *Avian Dis.* 2008;52:367-374.<https://doi.org/10.1637/8323-041808-Review.1>
 22. Gharaibeh S, Hailat A. *Mycoplasma gallisepticum* experimental infection and tissue distribution in chickens, sparrows and pigeons. *Avian Pathol.* 2011;40:349-354.<https://doi.org/10.1080/03079457.2011.582480>
 23. Ferguson-Noel N. Mycoplasmosis. In: Swayne DE, Glisson JR, McDougald LR, Nolan LK, Suarez DL, Nair VL, editors. *Diseases of Poultry.* 13th ed. Ames, IA: Blackwell-Wiley Publishing; 2013. p. 875-876.<https://doi.org/10.1002/9781119421481.ch21>
 24. Reinhardt AK, Kempf I, Kobisch M, Gautier-Bouchardon AV. Fluoroquinolone resistance in *Mycoplasma gallisepticum*: DNA gyrase as primary target of enrofloxacin and impact of mutations in topoisomerases on resistance level. *J Antimicrob Chemother.* 2002;50:589-592.<https://doi.org/10.1093/jac/dkf158>
 25. Grodio J, Buckles E, Schat K. Production of house finch (*Carpodacus mexicanus*) IgA specific anti-sera and its application in immunohistochemistry and in ELISA for detection of *Mycoplasma gallisepticum*-specific IgA. *Vet Immunol Immunopathol J.* 2009;132:288-294.<https://doi.org/10.1016/j.vetimm.2009.06.006>
 26. Gaunson E, Philip C, Browning G. Lymphocytic infiltration in the chicken trachea in response to *Mycoplasma gallisepticum* infection. *Microbiol J.* 2000;146:1223-1229.<https://doi.org/10.1099/00221287-146-5-1223>
 27. Gaunson E, Philip C, Whithear K, Browning G. The cellular immune response in the tracheal mucosa to *Mycoplasma gallisepticum* in vaccinated and unvaccinated chickens in the acute and chronic stages of disease. *Vaccine J.* 2006;24:2627-2633.<https://doi.org/10.1016/j.vaccine.2005.12.008>
 28. Majumder S. Role of *Mycoplasma gallisepticum* and host airway epithelial cell interaction in inflammation. [dissertation]. Storrs: University of Connecticut; 2014. Available from:<https://opencommons.uconn.edu/dissertations/651>
 29. Javed MA. Correlates of immune protection in chickens vaccinated with *Mycoplasma gallisepticum* strain GT5 following challenge with pathogenic *M. gallisepticum* strain R (low). *Infect*



- Immun. 2005;73(9):5410-5419.<https://doi.org/10.1128/IAI.73.9.5410-5419.2005>
30. Ley DH. Mycoplasma gallisepticum infection. In: Saif YM, Fadly AM, Glisson JR, McDougald LR, Nolan LK, Swayne DE, editors. Diseases of Poultry. 12th ed. Ames: Blackwell Publishing Professional; 2008. p. 807-834. Available from: <https://www.thepoultrysite.com/publications/diseases-of-poultry>
 31. Blanchard A, Browning G. Mycoplasmas: molecular biology pathogenicity and strategies for control. Boca Raton: CRC Press; 2005.
 32. Feberwee A, Von Banniseht-Wysmuller T, Vernooij J, Gielkens Stegeman A. The effect of vaccination with a bacterin on the horizontal transmission of Mycoplasma gallisepticum. Avian Pathol. 2006;35:35-37.<https://doi.org/10.1080/03079450500465700>
 33. Al-Momani W, Abu-Basha E, Janakat S, Nicholas RAJ, Ayling RD. In vitro antimycoplasmal activity of six Jordanian medicinal plants against three Mycoplasma species. Trop Anim Health Prod. 2007;39:515-519.<https://doi.org/10.1007/s11250-007-9033-1>
 34. Altamimi MKA, Al-Zubaidi MTS. High Prevalence of Cryptosporidium meleagridis in Domestic Pigeons (Columba livia domestica) Raises a Prospect of Zoonotic Transmission in Babylon Province, Iraq. The Third International Conference for Postgraduate Scientific Research, December 15, 2020 College of Veterinary Medicine, University of Baghdad. 2020;44(E0):7-13.[https://doi.org/10.30539/ijvm.v44i\(E0\).1012](https://doi.org/10.30539/ijvm.v44i(E0).1012)
 35. Ahmed AI. Molecular Characterization of Infectious Bursal Disease Virus Isolated from Naturally Infected Broiler Chickens in Erbil, Iraq. The Third International Conference for Postgraduate Scientific Research, December 15, 2020 College of Veterinary Medicine, University of Baghdad. 2020;44(E0):21-27.[https://doi.org/10.30539/ijvm.v44i\(E0\).1015](https://doi.org/10.30539/ijvm.v44i(E0).1015)
 36. Fadhil LT, Faraj AA, AL-Amery AM. Trichomonas gallinae Identification and Histopathological Study in Pigeon (Columba livia domestica) in Baghdad City, Iraq. The Third International Conference for Postgraduate Scientific Research, December 15, 2020 College of Veterinary Medicine, University of Baghdad. 2020;44(E0):57-63.[https://doi.org/10.30539/ijvm.v44i\(E0\).1022](https://doi.org/10.30539/ijvm.v44i(E0).1022)
 37. Naser R, Khaleel IM. The Arterial Vascularization of the Small and Large Intestine in Adult Male Turkeys (Meleagris gallopavo). The Third International Conference for Postgraduate Scientific Research, December 15, 2020 College of Veterinary Medicine, University of Baghdad. 2020;44(E0):69-74.[https://doi.org/10.30539/ijvm.v44i\(E0\).1024](https://doi.org/10.30539/ijvm.v44i(E0).1024)
 38. Ibrahim RM, Al-Rubaie HMA. Molecular Detection of Avian Malaria (Plasmodium gallinaceum) in Local Domesticated Breed Chickens (Gallus gallus domesticus) in Baghdad. The Third International Conference for Postgraduate Scientific Research, December 15, 2020 College of Veterinary Medicine, University of Baghdad. 2020;44(E0):75-79.[https://doi.org/10.30539/ijvm.v44i\(E0\).1025](https://doi.org/10.30539/ijvm.v44i(E0).1025)
 39. Ayman U, Jahid MA, Alam MR, Das SK. Morphohistology and Biometric Characteristics of Cecal Tonsils of Sonali Chicken at Post-Hatching Ages. Iraqi J Vet Med. 2021;45(2):1-6.<https://doi.org/10.30539/ijvm.v45i2.1254>
 40. Ali AJ. Isolation, identification and some aspects pathogenicity of Mycoplasma gallisepticum in broiler chickens. [dissertation]. Baghdad: College of Veterinary Medicine, University of Baghdad; 2019.
 41. Mohammed AK, Abd-Al-Shaheed DA. Minimum inhibition concentration of two antibiotics and two plants extract against Mycoplasma gallisepticum in Iraq. Int J Psychosocial Rehabil. 2021a;25(2):1189-1203.
 42. Mohammed AK, Abd-Al-Shaheed DA. Prevalence of Mycoplasma gallisepticum in poultry from Iraq. Online J Vet Res. 2021b;25(7):459-464.
 43. Mohammed AK. Laboratory Study of Mycoplasma gallisepticum on Pregnant Mice and Evaluate the Antimicrobial Activity against it. [dissertation]. Baghdad: College of Veterinary Medicine, University of Baghdad; 2022.